

## PRELIMINARY PHYTOCHEMICAL SCREENING AND IN-VITRO ANTIBACTERIAL ACTIVITY ON ASPARAGUS RACEMOSUS ROOT EXTRACT

K.RAVISHANKAR\*, G.V.N.KIRANMAYI<sup>1</sup>, T.MEHER LALITHA, T.PRIYANKA, T.RANJITH, S.B.V.SOMESWARAO, V.R.KRISHNAM RAJU and A.V.DIVYA

Sri Sai Aditya Institute of Pharmaceutical Sciences and Research, Adb Road, Surampalem, East Godavari District, Andhra Pradesh, India.

### ABSTRACT

The purpose of present work is to study medicinally active substances present in ethanol-extract obtained from roots of *Asparagus racemosus*. Preliminary Phytochemical screening of the extracts revealed the presence of Alkaloids, Carbohydrates, Glycosides, Phenolic compounds, tannins, Saponins, Steroids and Flavonoids. The presence of these bioactive constituents is associated with the antibacterial activity of the plant. The root extracts of *Asparagus racemosus* solvated by ethanol, showed the spectrum of inhibition on *Staphylococcus aureus*, *Bacillus subtilis*, *Staphylococcus wernerii*, *Pseudomonas putida*, *Pseudomonas aeruginosa* and *Proteus mirabilis* by Cylinder plate method. The observations revealed significant zone of inhibition and supports to antibacterial activity.

**Keywords:** *Asparagus racemosus*; Phytochemical; antibacterial activity.

### INTRODUCTION

Nature has been a source of medicinal agents since times immemorial. The importance of herbs in the management of human ailments cannot be over emphasized. It is clear that the plant kingdom harbors an inexhaustible source of active ingredients invaluable in the management of many intractable diseases. However, these complementary components give the plant as a whole a safety and efficiency much superior to that of its isolated and pure active components<sup>1</sup>. There are several reports on the antimicrobial activity of different herbal extracts in different regions of the world<sup>2-5</sup>. Because of the side effects and the resistance that pathogenic microorganisms

build against antibiotics, recently much attention has been paid to extracts and biologically active compounds isolated from plant species used in herbal medicine<sup>6</sup>. Approximately 20% of the plants found in the world have been submitted to pharmacological or biological test, and a substantial number of new antibiotics introduced on the market are obtained from natural or semi-synthetic resources<sup>7</sup>. In the present study, medicinal plant *Asparagus racemosus* belonging to the family Asparagaceae was selected to assess antibacterial potential.

*Asparagus racemosus* (Shatavari) is recommended in traditional medicine for the prevention and treatment of gastric

ulcers, dyspepsia, diarrhea, nervous disorders<sup>8</sup>. Besides use in the treatment of and dysentery, the plant also has antioxidant, immunostimulant, anti-dyspepsia and antitussive effects<sup>9</sup>. Scanty work has reported an antimicrobial activity of whole plant<sup>10</sup>. *Asparagus racemosus* have been used for the treatment of the ulcers, depression, inflammation cancer, lithiasis, Hepatotoxicity, diabetes<sup>11-30</sup>. Thus study is aimed to demonstrate and determine the antibacterial effects of *Asparagus racemosus* root on various strains of bacteria.

## MATERIALS AND METHODS

### Plant material

Fresh plant roots were collected from local area of East Godavari district and the plants were identified by Botanist Dr. T. U. Ragharam. The plant was dried in shade, and then pulverized into powder.

### Preparation of crude ethanol extract

The root powder was repeatedly macerated with 95% ethanol in a percolator. The combined filtrate was evaporated to dryness under reduced pressure at 40–50°C. The resulting crude ethanol extract was then stored at 10–15°C.

### Test Organism Used

The various organisms like *Staphylococcus aureus* ATCCBAA 1026, *Bacillus subtilis* ATCC 11774, *Staphylococcus wernerii* ATCC 27836, *Pseudomonas putida* ATCC 700007, *Pseudomonas aeruginosa* ATCC 10662, *Proteus mirabilis* ATCC 14153, *Escherichia coli* ATCC 10536, *Klebsella pneumonia* ATCC 33495 are procured from Microbes Speciality Lab Danavaipeta, Rajahmundry, East Godavari District 533103, Andhra Pradesh, India.

### Antimicrobial Agent

The reference standard Gentamycin was procured from Pradeep Organics and chemicals Pvt. Ltd, Hyderabad.

### Phytochemical screening

The powdered root was evaluated for qualitative determination of major phytoconstituents i.e. Alkaloids, Carbohydrates, Glycosides, Phenolic

compounds, tannins, Saponins, Steroids, Flavonoids; which were further confirmed by thin layer chromatography.

### Qualitative screening

Alkaloid detection was carried out by extracting 1 g powdered sample with 5 ml methanol and 5 ml of 2N HCl; and then treating the filtrate with Mayer's and Wagner's reagents. The samples were scored positive on the basis of Reddish brown or cream precipitation. Flavonoids were tested by heating 1 g powdered sample with 10 ml ethyl acetate over a steam bath (40–50°C) for 5 min; filtrate was treated with 1 ml dilute ammonia. A yellow coloration demonstrated positive test for Flavonoids. Saponins content was determined by boiling 1 g powdered sample in 10 ml distilled water for 15 min and after cooling, the extract was shaken vigorously to observe froth formation. Cardiac glycosides were identified by **Borntrager's test**. Ammonical layer turning to pink was indicative of cardenolides/cardiac glycosides<sup>31</sup>.

### Thin layer chromatography (TLC)

TLC plates were prepared by using silica gel G for TLC, were left overnight for air drying. These plates were activated by hot air oven at 100°C for 1hr. Cold alcoholic extract was plotted on TLC plates<sup>32</sup>. The plates were dried and developed in suitable solvents for rapid screening. Pure ethyl acetate, 50% chloroform/methanol, 1:1 ethyl acetate/methanol. The plates were run in the above solvent systems and dried at room temperature. Derivatisation of TLC plates was done by spraying 10% H<sub>2</sub>SO<sub>4</sub> in methanol. Different bands were observed and corresponding R<sub>f</sub> values are determined. R<sub>f</sub> value of each spot was calculated as

$$R_f = \frac{\text{Distance travelled by the solute}}{\text{Distance travelled by the solvent.}}$$

### Antibacterial Assay

Root extract of *Asparagus racemosus* was evaluated for antibacterial activity against several Gram Positive and Gram Negative organisms.

The antibacterial activity of ethanolic extract was performed using Agar cup-plate

method. 20ml of sterile nutrient agar medium was poured into sterile Petri dishes and allowed to solidify. The Petri dishes were incubated at 37°C for 24 hours to check for sterility. The medium was seeded with the organisms by pour plate method using sterile top agar (4 ml) contained 1 ml culture. Bores were made on the medium using sterile borer. Dried ethanolic extract of roots of *Asparagus racemosus* was dissolved in water to obtain different concentrations (100 , 300 and 500mg/ml) and sterilized by filtration through a Whatman filter paper no. 1, and 0.05 ml of the different concentrations of extract were added to the respective bores. 0.05ml of Gentamycin at a concentration of (25 µg/ml) was taken as standard reference. All the plates were kept in a

refrigerator at 2 to 8° c for a period of 2 hours for effective diffusion of test compounds and standards. Later, they were incubated at 37°C for 24 hours. The presence of definite zone of inhibition of any size around the cup indicated antibacterial activity. The diameter of the zone of inhibition was measured and recorded.

## RESULTS AND DISCUSSION

### Preliminary Phytochemical screening

Phytochemical screening of the extracts of *Asparagus racemosus* revealed, the presence of Alkaloids, Carbohydrates, Glycosides, Phenolic compounds, tannins, Saponins, Steroids, and Flavonoids. (Table 1).

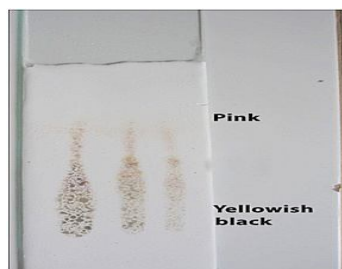
**Table1: Preliminary Phytochemical analysis of *Asparagus racemosus* root extract**

Components	<i>Asparagus racemosus</i> (ethanolic root extract)
Alkaloids	+
Carbohydrates	+
Glycosides	+
Phenolic compounds and tannins	+
Proteins and amino acids	-
Saponins	+
Steroids	+
Flavonoids	+

### Thin layer chromatography

The presence of phytoconstituents was further confirmed by thin layer chromatography and their  $R_f$  values have been presented as 0.7 (Figure 1). The

components were best resolved in screening system using pure ethyl acetate, 50% chloroform/methanol, 1:1 ethyl acetate/methanol.



**Yellow Colour** Indicates Presence of Tannins, Flavonoids.

**Pink Colour** Indicates Presence of Steroids.

**Fig. 1: TLC of *Asparagus racemosus***

### Antibacterial Activity

The root extract of *Asparagus racemosus* was studied for antibacterial activity

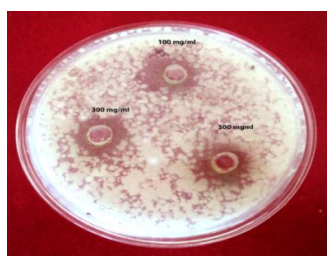
employing standard cylinder method. Microbes used were *Bacillus subtilis*, *Staphylococcus aureus*, *Staphylococcus wernerii*, *Pseudomonas aeruginosa* and

*Escherichia coli*, *Proteus mirabilis*, *Klebsiella pneumoniae*, *Pseudomonas putida*. Both gram-positive and gram-negative bacteria were sensitive to the extract. The zone of inhibition recorded for various organisms was found *Staphylococcus aureus* (18mm), *Bacillus subtilis* (13mm), *Staphylococcus wernerii* (14mm), *Pseudomonas putida* (17mm), *Pseudomonas aeruginosa* (15mm),

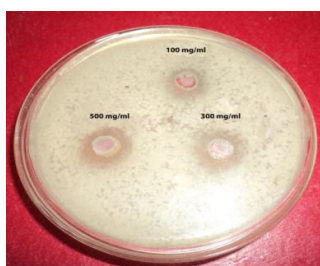
*Proteus mirabilis* (15mm). Activity of ethanolic extract of the plant was comparable to that of Reference Standard drug Gentamycin disc (25µg). *Asparagus racemosus* root extract exhibited good antimicrobial activity and results were tabulated along with figures (Table 2; figure 2,3&4).

**Table 2: Antibacterial activity of *Asparagus racemosus* root extract**

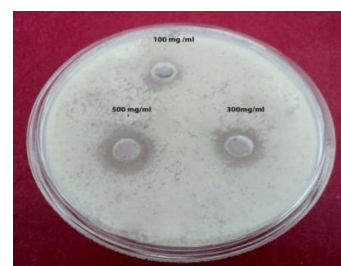
Microorganism	Zone of inhibition(mm)			
	100mg/ml	300mg/ml	500mg/ml	Gentamycin 25µg/ml
<b>Gram positive</b>				
<i>S.aureus</i>	16 ± 0.21	17 ± 0.36	18 ± 0.15	12 ± 0.12
<i>B.subtilis</i>	10 ± 0.62	12 ± 0.69	13 ± 0.36	14 ± 0.32
<i>S.wernerii</i>	12 ± 0.48	14 ± 0.48	14 ± 0.52	13 ± 0.28
<b>Gram negative</b>				
<i>P.putida</i>	13 ± 0.26	13 ± 0.52	17 ± 0.25	15 ± 0.21
<i>P.aeruginosa</i>	12 ± 0.36	13 ± 0.39	15 ± 0.53	16 ± 0.12
<i>Proteus mirabilis</i>	13 ± 0.14	14 ± 0.28	15 ± 0.63	13 ± 0.36
<i>Escherichia coli</i>	8±0.39	9±0.35	9±0.24	13±0.28
<i>Klebsiella pneumoniae</i>	7±0.16	8±0.26	9±0.52	14±0.32



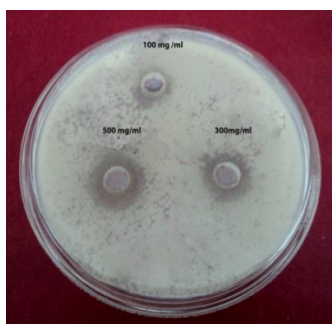
**Staphylococcus aureus**



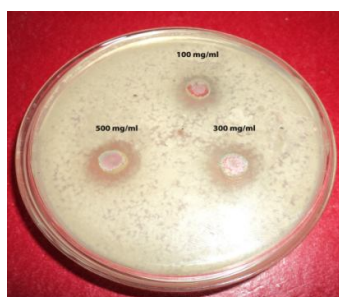
**Bacillus subtilis**



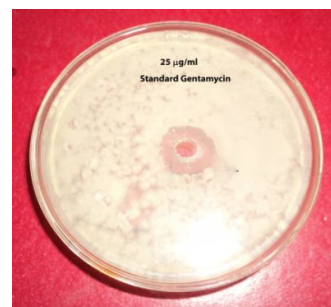
**Staphylococcus wernerii**



**Pseudomonas aeruginosa**



**Pseudomonas putida**



**standard**

**Fig. 2: Zone Of Inhibition of *Asparagus racemosus* root extract**

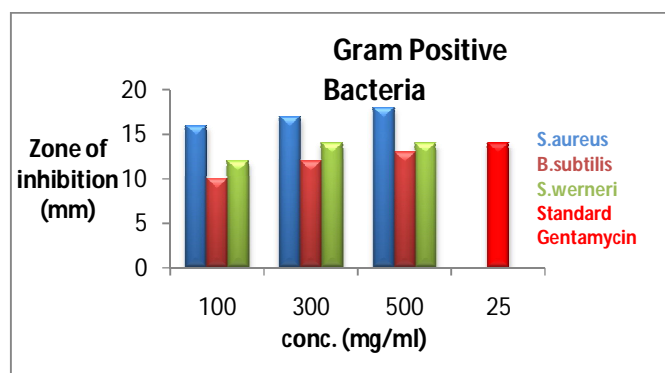


Fig. 3: Inhibition Zone of *Asparagus racemosus* Root Extract against Gram Positive Organisms

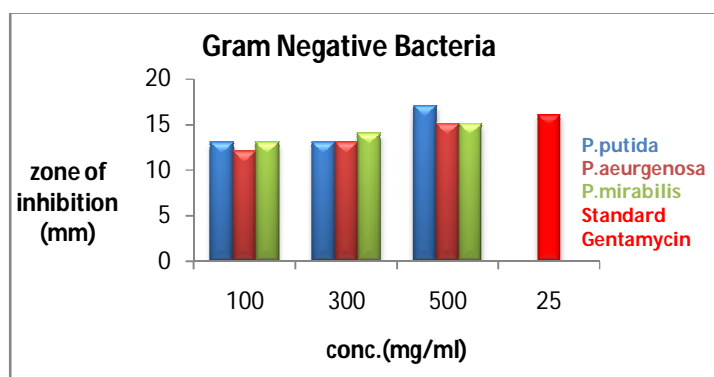


Fig. 4: Inhibition Zone of *Asparagus racemosus* Root Extract against Gram negative Organisms

## CONCLUSION

The scientific paper establishes that *Asparagus racemosus* root extract has good significant antibacterial activity against *Staphylococcus aureus*, *Bacillus subtilis*, *Staphylococcus weneri*, *Pseudomonas putida*, *Pseudomonas aeruginosa*, *Proteus mirabilis*. The antibacterial activity of root extract with different concentrations 100, 300 and 500 mg/ml was very well compared with standard reference drug Gentamycin 25 µg/ml. The maximum zone of inhibition with root extract of *Asparagus racemosus* at various concentrations like 100, 300 and 500 mg/ml are 16, 17 and 18 mm. Further investigations can be carried out in order to isolate new compounds from the plant root extract and to evaluate the bioactivities as it is necessary to introduce new biologically safe phytochemical compounds which are necessary to suppress the growth of the microorganisms.

## REFERENCES

1. Shariff ZU. Chemical composition and antimicrobial activity of the essential oils from the gum of Turkish Pistachio (*Pistacia vera* L.) J Agric Food Chem. 2001;52(12): 3911 – 3914.
2. Chung PY, Chung LY and Ngeow YF. Antimicrobial activities of Malaysian plant species. Pharm Bio. 2004;42:292–300.
3. Nair R and Chanda SV. Antibacterial activity of some medicinal plants of Saurashtra region. J Jissue Res. 2004;4:117–120.
4. De N and Ifeoma E. Antimicrobial effects of components of the bark extracts of Neem. J Technol. Dev. 2002;8:23–28.
5. Nair R, Kalariya T and Chanda S. Antibacterial activity of some



- selected Indian medicinal flora. Turk J Biol. 2005;29:41-47.
6. Essawi T and Srour M. Screening of some palestnian medicinal plants for antibacterial activity. J Ethnopharmacol. 2000;70:343-349.
  7. Mothana RA and Lindequist U. Antimicrobial activity of some medicinal plants of the island soqotra. J Ethnopharmacol. 2005;96:177-181.
  8. Goyal RK, Singh J and Lal H. Asparagus racemosus Indian Journal of Medical Sciences. 2003;57(9):408-14.
  9. Bopana N and Saxena S. Asparagus racemosus ethnopharmacological evaluation and conservation needs. Journal of Ethnopharmacology. 2007;110(1):1-15.
  10. Mandal SC, Nandy A, Pal M and Saha BP. Evaluation of antibacterial activity of Asparagus racemosus wild root. Phytotherapy Research. 2000;14(2):118-9.
  11. Bhatnagar M and Sisodia SS. Antisecretory and antiulcer activity of Asparagus racemosus Willd. against indomethacin plus pyloric ligation-induced gastric ulcer in rats. Journal of Herbal Pharmacotherapy. 2006;6(1):13-20.
  12. Sairam K, Priyambada S, Aryya NC and Goel RK. Gastroduodenal ulcer protective activity of Asparagus racemosus: an experimental, biochemical and histological study. Journal of Ethnopharmacology. 2003;86(1):1-10.
  13. Garabadu D, Muruganandam AV, Joshi VK and Krishnamurthy S. Antidepressant activity of Asparagus racemosus in rodent models. Singh GK. Pharmacology, Biochemistry & Behavior. 2009;91(3):283-90.
  14. Lee do Y, Choo BK, Yoon T, Cheon MS, Lee HW, Lee AY and Kim HK. Anti-inflammatory effects of Asparagus cochinchinensis extract in acute and chronic cutaneous inflammation. Journal of Ethnopharmacology. 2009;121(1):28-34.
  15. Liu W, Huang XF, Qi Q, Dai QS, Yang L, Nie FF, Lu N, Gong DD, Kong LY and Guo QL. Asparanin A induces G(2)/M cell cycle arrest and apoptosis in human hepatocellular carcinoma HepG2 cells. Biochemical & Biophysical Research Communications. 2009;381(4):700-5.
  16. Visavadiya NP, Soni B and Madamwar D. Suppression of reactive oxygen species and nitric oxide by Asparagus racemosus root extract using in vitro studies. Cellular & Molecular Biology. 2009;55:1083-95.
  17. Wiboonpun N, Phuwapraisirisan P and Tip-pyang S. Identification of antioxidant compound from Asparagus racemosus Phytotherapy Research. 2004;18(9):771-3.
  18. Kamat JP, Boloor KK, Devasagayam TP and Venkatachalam SR. Antioxidant properties of Asparagus racemosus against damage induced by gamma-radiation in rat liver mitochondria. Journal of Ethnopharmacology. 2000;71(3):425-35.
  19. Mandal SC, Kumar CKA, Mohana Lakshmi S, Sinha S, Murugesan T and Saha BP. Antitussive effect of Asparagus racemosus root against sulfur dioxide-induced cough in mice. Pal M Fitoterapia. 2000;71(6):686-9.
  20. Ojha R, Sahu AN, Muruganandam AV, Singh GK and Krishnamurthy S. Asparagus racemosus enhances memory and protects against amnesia in rodent models. Brain & Cognition. 2010;74(1):1-9.
  21. Christina AJ, Ashok K, Packialakshmi M, Tobin GC, Preethi J and Muruges N. Antilithiatic effect of Asparagus racemosus Willd on ethylene glycol-induced lithiasis in male albino Wistar rats. Methods & Findings in Experimental & Clinical Pharmacology. 2005;27(9):633-8.
  22. Zhu X, Zhang W, Zhao J, Wang J and Qu W. Hypolipidaemic and hepatoprotective effects of ethanolic and aqueous extracts from

- Asparagus officinalis L. by-products in mice fed a high-fat diet. *Journal of the Science of Food & Agriculture*. 2010;90(7):1129-35.
23. Thakur M, Chauhan NS, Bhargava S and Dixit VK. A comparative study on aphrodisiac activity of some ayurvedic herbs in male albino rats. *Archives of Sexual Behavior*. 2009;38(6):1009-15.
  24. Prabha T, Kumar MM, Dorababu M, Prakash Singh G and Goel RK. Teratogenicity of Asparagus racemosus Willd root a herbal medicine. *Indian Journal of Experimental Biology*. 2006;44(7):570-3.
  25. Gautam M, Saha S, Bani S, Kaul A, Mishra S, Patil D, Satti NK, Suri KA, Gairola S, Suresh K, Jadhav S, Qazi GN and Patwardhan B. Immunomodulatory activity of Asparaguracemosus on systemic Th1/Th2 immunity: implications for immunoadjuvant potential. *Journal of Ethnopharmacology*. 2009;121(2):241-7.
  26. Gautam M, Diwanay S, Gairola S, Shinde Y, Patki P and Patwardhan B. Immunoadjuvant potential of Asparagus racemosus aqueous extract in experimental system. *Journal of Ethnopharmacology*. 2004;91(2-3):251-5.
  27. Pandey SK, Sahay A, Pandey RS and Tripathi YB. Effect of Asparagus racemosus rhizome (Shatavari) on mammary gland and genital organs of pregnant rat. *Phytotherapy Research*. 2005;19(8):721-4.
  28. Parihar MS and Hemnani T. Experimental excitotoxicity provokes oxidative damage in mice brain and attenuation by extract of Asparagus racemosus. *Journal of Neural Transmission*. 2004;111(1):1-12.
  29. Agrawal A, Sharma M, Rai SK, Singh B, Tiwari M and Chandra R. The effect of the aqueous extract of the roots of Asparagus racemosus on hepatocarcinogenesis initiated by diethylnitrosamine.. *Phytotherapy Research*. 2008;22(9):1175-82.
  30. Hannan JM, Marenah L, Ali L, Rokeya B, Flatt PR and Abdel-Wahab YH. Insulin secretory actions of extracts of Asparagus racemosus root in perfused pancreas, isolated islets and clonal pancreatic beta-cells. *Journal of Endocrinology*. 2007;192(1):159-68.
  31. Harborne JB. *Phytochemical methods – A guide to modern techniques of plant analysis*. 3rd edition. New Delhi: Springer Pvt. Ltd; 2005.
  32. Wagner H and Bladt S: *Plant drug analysis-A thin layer chromatography atlas*. 2nd edition. New Delhi: Thompson Press Ltd; 2004.