

INVITRO ANTICANCER ACTIVITY OF *TECOMASTANS* (L.) ETHANOLIC LEAF EXTRACT ON HUMAN BREAST CANCER CELL LINE (MCF-7)

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ABSTRACT

Tecomastans (L.) (Family: Bignoniaceae) is called as yellow elder in English. Traditionally flowers and bark are used for treatment of various cancers. In current study crude ethanolic leaf extract of *Tecomastans* (L.) leaves were examined for their anticancer activity. To determine *invitro* anticancer activity, different concentrations of crude extract were tested on MCF-7 cancer cell line by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. *Tecomastans* leaf extract showed a significant antiproliferative activity and a dose dependent effect was observed. Minimum inhibition of 14.6% was shown by extract at concentration 7.8 µg/ml and maximum inhibition (95.9%) was observed at 1000 µg/ml. The plant extract showed activity in potential range for further investigation on cancer cells.

Keywords: *Tecomastans*, MCF-7 cancer cell line, MTT assay, anticancer activity.

INTRODUCTION

Cancer is the second leading cause of death all over the world¹ is a multifactorial, multifaceted and multimechanistic disease requiring a multidimensional approach for its treatment, control and prevention². The major causes of cancer are smoking, dietary imbalances, hormones and chronic infections leading to chronic inflammation³. Age is also a primary risk factor for most cancers, with about 77% of all cancers diagnosed among people aged 55 or older⁴. Breast cancer is the most common form of cancer in women worldwide⁵. According to an estimate, 50% of breast cancer and 37% of prostate cancer patients use herbal products⁶. Several chemopreventive agents are used to treat cancer, but they cause toxicity that prevents their usage⁷. In this context, the natural

products derived from medicinal plants have gained significance in the treatment of cancer⁸.

Tecomastans (L.) belongs to the family Bignoniaceae are distributed worldwide, mostly occur in tropical and sub tropical countries. However a number of temperate species also grow in North America and East Asia⁹. *T.stans* is a shrub or small tree, 5-7.6 m in height. Bark is pale brown to grey and roughens with age. Leaves are compound and imparipinnate with 2 to 5 pairs of leaflets and a larger single terminal leaflet. Leaflets are lanceolate, up to 10 cm long, with serrated margins, mid-green above and soft to the touch. Flowers occur in clusters at the ends of the branches and are trumpet shaped with 5 rounded lobes, 6 cm long, pale to bright yellow, with faint orange stripes at the throat.

Fruits are narrow, slightly flattened to pointed capsules, up to 20 cm long, containing many winged seeds; green when young, pale brown on ripening and remain on the tree in untidy clusters for many months¹⁰. Traditional use of leaves of *T.stans* in throughout Mexico and central America for diabetes and urinary disorder control^{11,12,13}. Roots are used as diuretic and vermifuge¹⁴. Traditionally flowers and bark are used for treatment of various cancers. The stem barks showed better antimicrobial activity¹⁵. Its Leaf shows Anthelmintic Activity¹⁶, Antispasmodic effect¹⁷, Antibacterial activity¹⁸, Anticancer Activity¹⁹, Wound Healing property²⁰. Flower shows Antidiabetic Activity²¹ & anticancer activity²² while roots shows Antibacterial Acitivity²³. Aerial Parts shows Antioxidant Activity²⁴. Bark shows Wound Healing property²⁵.

MATERIALS AND METHODS

Plant Collection and Extraction

The leaves of *Tecomastans* were collected from local area of Avadi (west Chennai) in the month of march. The plant was identified and authenticated by Dr. P. Jayaraman (PARC, chennai), bearing a voucher Reg.no of PARC/2012/1141. The plant material was air dried at room temperature, coarsely powdered and stored in air tight container and used for further extraction. The dried powder (50gm) was extracted successively with ethanol (60°C) by using a Soxhlet apparatus for 8 hrs.

Phytochemical screening

Qualitative chemical tests were carried out using extracts from plant to identify the phytochemicals²⁶.

Cell line and Culture

Breast cancer- MCF-7 cell lines was obtained from National centre for cell sciences, Pune (NCCS). The cells were maintained in Minimal Essential Media (MEM) supplemented with

10% Fetal bovine serum (FBS), penicillin (100 U/ml), and streptomycin (100µg/ml) in a humidified atmosphere of 50µg/ml CO₂ at 37 °C.

Reagents

MEM was purchased from Hi Media Laboratories FBS was purchased from Cistron laboratories Trypsin, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl- tetrazolium bromide (MTT) and Dimethyl Sulfoxide (DMSO) were purchased from (Sisco research laboratory chemicals, Mumbai). All of other chemicals and reagents were obtained from Sigma Aldrich, Mumbai.

In vitro assay for Cytotoxicity activity (MTT assay)

The Cytotoxicity of samples on MCF-7 was determined by the MTT assay²⁷. Cells (1 × 10⁵/well) were plated in 100µl of medium/well in 96-well plates (Costar Corning, Rochester, NY). After 48 hours incubation the cell reaches the confluence. Then, cells were incubated in the presence of various concentrations of the samples in 0.1% DMSO for 48h at 37°C. After removal of the sample solution and washing with phosphate-buffered saline (pH 7.4), 20µl/well (5mg/ml) of 0.5% 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl--tetrazolium bromide (MTT) phosphate- buffered saline solution was added. After 4h incubation, 0.04M HCl/ isopropanol was added. Viable cells were determined by the absorbance at 570nm. Measurements were performed and the concentration required for alinhibition Concentration (IC50) was determined graphically. The absorbance at 570nm was measured with a UV- Spectrophotometer using wells without sample containing cells as blanks. The effect of the samples on the proliferation of MCF-7 was expressed as the % cell viability & % Cell death using the following formulas:

$$\% \text{ cell viability} = A_{570} \text{ of treated cells} / A_{570} \text{ of control cells} \times 100\%.$$

$$\% \text{ Cell death} = (\text{Control OD} - \text{Sample OD}) / \text{Control OD} \times 100.$$

RESULTS

Phytochemical screening reveals the presence of carbohydrates, proteins, saponins, flavanoids, alkaloids, tannins, phenolic compounds.

Preliminary reports have attributed the leaves of *T. stans* with *in vitro* Anticancer activity (Table.1, Fig.1 & 2). The Photomicrograph of MCF-7 cell line at various concentrations are shown in Fig.3. The IC₅₀ was found to be 64.5µg/ml. Accumulating evidence clearly indicates that apoptosis is a critical molecular target by dietary bioactive agents for the prevention of cancer.

DISCUSSION

Plants are storehouse of "pre-synthesized" molecules that act as lead structures, which can be optimized for new drug development. In practice, a large number of cancer chemotherapeutic agents that are currently available in the market can be traced back to their plant source. Plant derived compounds;

in particular have gained importance in anticancer therapy and some of the new chemotherapeutic agents currently available for use includes paclitaxel, vincristine, podophyllotoxin and camptothecin, a natural product precursor from water soluble derivatives. Obviously natural products are extremely an important source of medicinal agents. Although there are some new approaches to drug discovery, such as combinatorial chemistry and computer based molecular modellingdesign, none of them can replace the importance of natural products in drug discovery and development^{28,29}. Literature data prove that triterpenes and flavonoids are biologically active against different strains of bacteria and many human cancer cell lines^{30,31,32}. The present study shows a dramatic *invitro* anticancer activity of Ethanolic Leaf extract of *Tecomastans*(L.) on human breast cancer cell line (MCF-7) at increasing concentrations. Inhibitory concentration (IC₅₀) was found to be 64.5µg/ml.

Table 1: *In vitro* Anticancer effect of *Tecomastans* leaf extract on MCF-7 cell line

S. No.	Concentration (µg/ml)	Dilutions	Absorbance (O.D)	Cell viability (%)	Cell death (%)
1	1000	Neat	0.02	4.1	95.9
2	500	1:1	0.09	18.7	81.3
3	250	1:2	0.13	27.3	72.7
4	125	1:4	0.19	39.5	60.5
5	62.5	1:8	0.24	50.0	50.0
6	31.2	1:16	0.29	60.4	39.6
7	15.6	1:32	0.36	75.0	25.0
8	7.8	1:64	0.41	85.4	14.6
9	Cell control	-	0.48	100	0

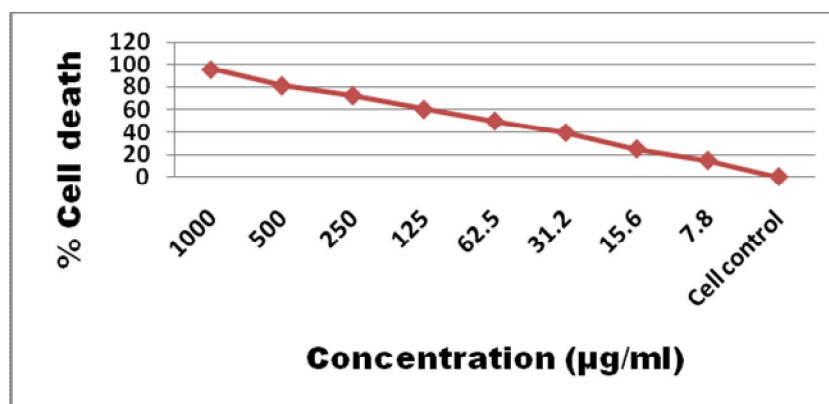


Fig. 1: Effect of ethanolic extract of *Tecomastans* leaf extract on MCF-7 cell death

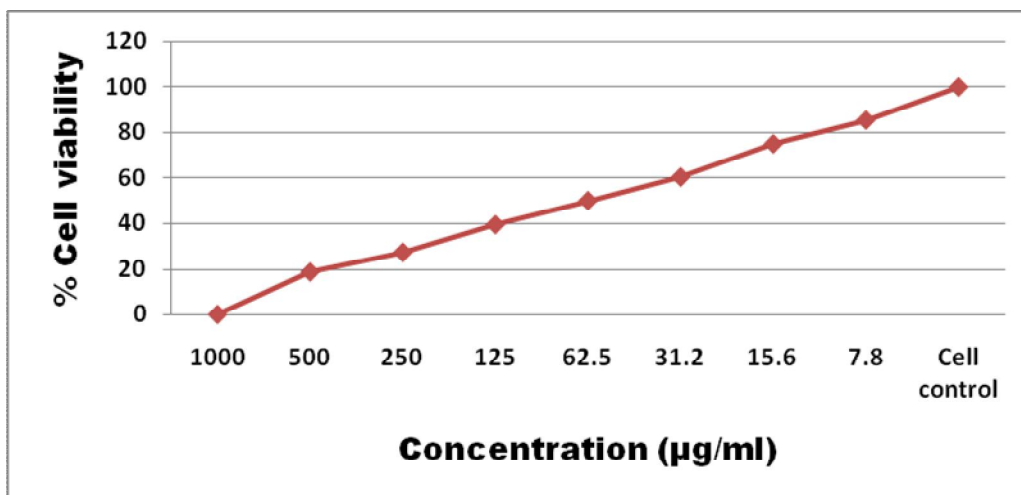


Fig. 2: Effect of ethanolic extract of *Tecomastans leaf extract* on MCF-7 cell viability

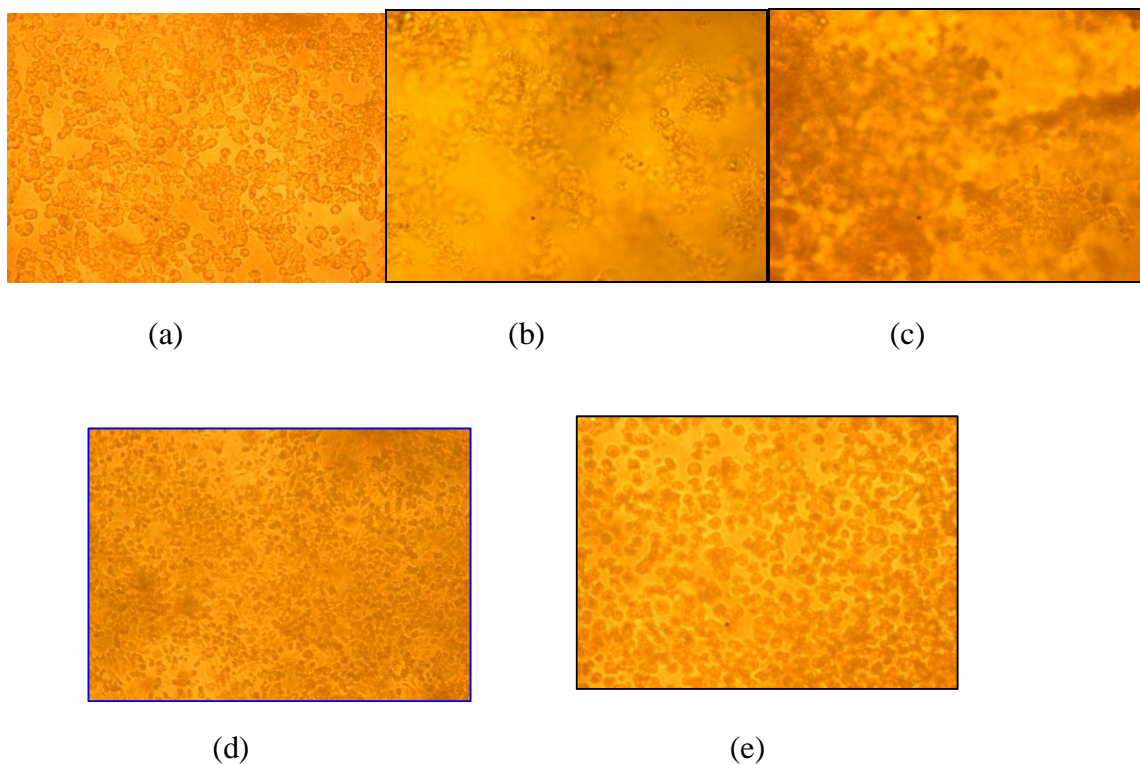


Fig. 3: Photomicrograph of MCF-7 cell line a) Control, b) Cell Toxicity at 1000µg/ml , c) Cell Toxicity at 250µg/ml, d) Cell Toxicity at 62.5µg/ml, e) Cell Toxicity at 31.2 µg/ml

CONCLUSION

The present results showed that *Tecomastans*(L.) ethanolic Leaf extract might be a potential alternative agent for human breast cancer therapy. Hence, it is anticipated that *T. stans* would be a useful pharmaceutical material to treat breast cancer. Future research should focus on the molecular mechanism of *T. stans* for anticancer action. There is a need for further investigation of this plant in order to identify and isolate its active anticancer principle(s) to treat breast cancer.

REFERENCES

1. Madhusudan S, Middleton MR. The emerging role of DNA repair proteins as predictive, prognostic and therapeutic targets in cancer. *Cancer Treatment Reviews*. 2005; 31(8): 603–617.
2. Jemal A, Murray T, Samuels A, Ghafoor A, Ward E, Thun M. cancer statistics. *Cancer J Clin*. 2003; 53:5-26.
3. Ames BN, Gold LS, Willett WC. The causes and prevention of cancer, *Proc. Natl. Acad. Sci. USA*. 1995; 92: 5258–5265.
4. National Cancer Institute (NCI). Cancer Control and Population Sciences: Research Findings [on-line]. Available at: <http://dccps.nci.nih.gov/ocs/prevalence/index.html>; 2003a.
5. Koduru S, Grierson DS, Afolayan AJ. Ethnobotanical information of medicinal plants used for treatment of cancer in the Eastern Cape Province. *South Africa. Curr. Sci*. 2007; 92: 906–908.
6. Richardson MA. Biopharmacologic and herbal therapies for cancer: research update from NCCAM. *J. Nut*. 2001; 131:3037S–3040S.
7. Kathiresan K, Boopathy NS, Kavitha S. Coastal vegetation – an underexplored source of anticancer drugs. *Nat. Prod. Rad*. 2006; 5: 115–119.
8. Gurib-Fakim A. Medicinal plants: traditions of yesterday and drugs of tomorrow. *Molecular Aspects of Medicine*. 2006; 27(1): 1–93.
9. Mohammed Rahmatullah, Walled Samarra, Rowan Jahan, Shahnaz Rahman, Nasima Sharmin ZUM, Emdad Ullah Miajee, Majeedul H, Chowdhury, Sazzadul Bari, Farhana Jamal, Anwarul Bashar ABM, Azad AK, Shamima Ahsan. An Ethnomedicinal, Pharmacological and Phytochemical Review of Some Bignoniaceae Family Plants and a Description of Bignoniaceae Plants in Folk Medicinal Uses in Bangladesh. *Advances in Natural and Applied Sciences*. 2010; 4(3): 236-53.
10. Orwa C, Mutua A, Kindt R, Jamnadass R, Simons A. *Agroforestry Database: a tree reference and selection guide version 4.0* (<http://www.worldagroforestry.org/af/treedb/>)
11. Winkelmann M. Frequently used medicinal plants in Baja California Norte. *Journal of Ethnopharmacology*. 1986; 18:109–31.
12. Lozoya X, Aguilar A, Camacho JR. Encuestas sobre el uso actual de las plantas en la Medicina Tradicional Mexicana. *Revista Médica del Instituto Mexicano del Seguro Social*. 1987; 25: 283–91.
13. Shapiro K, Gong WC. Natural products used for diabetes, *Journal of the American Pharmaceutical Association*. 2002; 42: 217–26.
14. Khare CP. *Indian Medicinal Plants An Illustrated Dictionary*. New Delhi; Springer, India. 2007: 648-649p.
15. Binuti OA, Lajubutu BA. Antimicrobial potentials of some plant species of the Bignoniaceae family. *African Journal of Medicine Science*. 1994; 23: 269-273.
16. Kumanan R, Sridhar C, Jayaveera KN, Sudha S, Rubesh Kumar S, Duganath N. Comparative Study of Anthelmintic Activity of Different Leaf Extracts of

- Tecomastans*(L.) on Adult Indian Earthworms. International Journal of Pharmaceutical and Clinical Research. 2010; 2(2): 63-65
17. GharibNaseri MK, AsadiMoghaddam M, Bahadoram S. Antispasmodic effect of *Tecomastans*(L.) Juss leaf extract on rat ileum, DARU. 2007; 15(3): 123-128.
 18. Senthilkumar CS, Suresh Kumar M, RajasekaraPandian M. In Vitro Antibacterial Activity Of Crude Leaf Extracts From *TecomaStans* (L) Juss. Et Kunth, *Coleus Forskohlii*And *Pogostemon Patchouli* Against Human Pathogenic Bacteria. Int.J. PharmTech Res. 2010; 2(1):438-442.
 19. PranayDogra. Study of Antibacterial and Anticancer Activity of Selected Trifoliolate Plants .biofrontiers. 2009; 1(2): 4-8.
 20. Das C, Dash S, Sahoo DC, Mohanty A. Evaluation OfMethanolic Bark Extract Of *TecomaStans*Linn, For Wound Healing In Albino Rats. Ijpt. 2010; 2(3): 735-742.
 21. Dhaked U, Gupta V, Singh DP, Nama G. antidiabetic activity of *tecomastans*flower. Pharmacologyonline. 2011; 1: 553-558.
 22. Kameshwaran S, Suresh V, Arunachalam G, Kanthal SK, Mohanraj M. *invitro*and *invivo*anticancer activity of methanolic extract of *tecomastans* flowers. IRJP. 2012; 3(3): 246-251.
 23. Ramesh T, Anusha V, Ravi Kumar A. Antibacterial AcitivityOfMethanolicExtract Of Roots Of *TecomaStans*. Int. J. Chem. Sci. 2009; 7(1): 6-8.
 24. Rasika C Torane, Gayatri S Kamble, Vaishali B Adsul, Chandrakant D Shendkar, Nirmala R Deshpande. Antioxidant Activity of Aerial Parts of *Tecomastans*. International Journal of Chemical and Analytical Science. 2011; 2(8): 130-132.
 25. Das C, Dash S, Sahoo DC, Mohanty A. Evaluation OfMethanolic Bark Extract Of *TecomaStans*Linn, For Wound Healing In Albino Rats. Ijpt. 2010; 2(3): 735-742.
 26. handelwal KR. Practical pharmacognosy, techniques and experiment, Nirali prakashan, pune, India. 2006; 16th edition.
 27. Mosmann T. Rapid colorimetric assay for cellular grow and survival: application to proliferation and cytotoxicity assays. J Immunol Meth. 1983;65: 55-63.
 28. Jagetia GC, Rao SK. Evaluation of Antineoplastic Activity *Guduchi* (*Tinosporacordifolia*) in Ehrlich Ascites Carcinoma bearing, Mice. Biol. Pharm. Bull. 2006; 29: 460-466.
 29. Heinrich M, Bremner P. Ethanobotany and ethanopharmacy-their role for anti-cancer drug development. Curr Drug Targets. 2006; 7: 239.
 30. Havsteen BH. The biochemistry and medical significance of the flavonoids. Pharmacol. Therap. 2002; 96: 67-202.
 31. Mahato SB, Nandy AK, Ray G. Triterpenoids. Phytochemistry. 1992; 31: 2199-2250.
 32. Min BS, Kim YH, Lee SM, Jung HJ, Lee JS, NA MK, Lee CO, Lee JP, Bae K. Cytotoxic triterpenes from *Crataeguspinnatifida*. Arch. Pharm. Res. 2000; 23:155-158.