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Research Article

APPLICATION OF A RP-HPLC METHOD FOR COMPARATIVE STUDY ON

THE DEGRADATION BEHAVIOR OF TWO ANGIOTENSIN II RECEPTOR

ANTAGONISTS, VALSARTAN AND LOSARTAN POTASSIUM

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ABSTRACT

Valsartan (VAL) and losartan potassium (LOS) are antihypertensive drugs, which are closely related in structure. Both drugs were subjected to stress conditions of hydrolysis (in acidic, neutral and alkaline media), oxidation (using hydrogen peroxide), dry heat and air. A reversed phase high performance liquid chromatographic method (RP-HPLC) was established by using acetonitrile: phosphate buffer (0.02M, pH 3.5), (60: 40 v/v) as a mobile phase at flow rate of 1.5 ml/ min and detection at 225 nm. Several factors were studied under the different experimental stress conditions. These factors include reagents concentrations, the applied temperature during stress testing and time of reaction. The degradation studies indicated that LOS is highly susceptible to oxidation, while VAL is very sensitive to acid hydrolysis and exhibits higher sensitivity to alkaline medium than LOS. The two drugs showed nearly the same stability under neutral hydrolysis. The RP-HPLC method was validated according to the ICH guidelines; the linearity range is 8-120 and 10-100 µg/mL for VAL and LOS, respectively; precision, accuracy, robustness, ruggedness, specificity were also evaluated. Thermal analysis of the two drugs was achieved in hot air oven and results show that valsartan is less thermally stable than losartan. The proposed RP-HPLC method is simple, accurate, reproducible, stability-indicating and suitable for routine determination of VAL or LOS in bulk and dosage forms.

Keywords: RP-HPLC; Stability indicating method; thermogravemetric; Valsartan.

INTRODUCTION

Valsartan (VAL) is designated as N-(1oxopentyl)-N-{[2'-(1H-tetrazol-5-yl)[1,1'-biphenyl]-4-yl]methyl}-L-valine2 (Fig.1a). losartan potassium (LOS), a potassium salt of : (2-butyl-4-chloro-1-{[2'-(1H-tetrazol-5yl)biphenyl-4-yl]methyl}-1H-imidazol-5-yl) methanol (Fig.1b). Valsartan and losartan are non-peptide, orally active angiotensin II (Type AT₁) receptor antagonists employed in the management of essential hypertension¹. Various methods have been reported for analysis of valsartan in pharmaceutical formulations alone or with other drugs in combinations, they include HPLC²⁻⁶, HPTLC⁷, capillary zone electrophoresis⁸, ⁹ spectrophotometric¹⁰⁻¹². Many analytical methods were reported for the

analysis of losartan especially in combination with other drugs, they include HPLC¹³⁻²¹, HPTLC²², electrophoretic^{23,24} and spectrophotometric²⁵⁻²⁸ methods.

Forced degradation plays a key role not just in the development of stability-indicating methods, but also in providing useful information about the degradation pathways and degradation products that could form during manufacture and storage. The information thus obtained will facilitate pharmaceutical development in areas such as manufacturing, formulation development, synthesis of degradation products (needed for purity determination) and packaging. Thus, stability testing is used to improve the quality of drug product. In the pre-clinical formulation stage, the selection of a stable drug product formula is the primary goal. This led us to study and compare the behavior of the two drugs valsartan and losartan in different mediums and conditions, using simple, sensitive, accurate and rapid RP-HPLC method.



Fig. 1a: Chemical structure of valsartan



potassium

EXPERIMENTAL MATERIALS

valsartan powder and its capsules, Disartan labeled to contain 80mg VAL per capsule were supplied from Global Napi (Egypt).

losartan powder and tablets, Kanzar labeled to contain 100 mg LOS per tablet were provided by Hikma company, Cairo- Egypt.

Acetonitrile and methanol, HPLC grade (rankem, India). All other chemicals are of analytical reagent grade. High pure water was prepared by using Millipore purification system.

Instrumentation and chromatographic conditions

The LC system, used for method development, forced degradation studies, and method validation was Agilent 1200 series liquid chromatographic RRHT (Rapid Resolution High Throughput) system comprising of binary pumps, column oven, autosampler and UV detector. The chromatographic column used was ACE C18 (250mm X 4.6mm X 5 μ). The mobile phase consists of a mixture of 0.02 mM sodium dihydrogen orthophosphate (pH adjusted to 3.5 using orthophosphoric acid) and acetonitrile in 40:60 ratio. The mobile phase was pumped from the solvent reservoir to the

column at a flow rate of 1.5 ml/min for 20 min. The column temperature was maintained at 40 °C. The eluate was monitored at 225 nm using UV detector and the injection volume was 20 µl. A shimadzu 1601 spectrophotometer with quartz cells of 1- cm optical path length and a Hanna Microprocessor HI 9321 with a combined glass-saturated calomel electrode were used.

Preparation of standard solutions

Standard stock solution (1mg/ml) of drug was prepared by dissolving 25 mg of valsartan or losartan in 5ml methanol or water, respectively. The solution was then ade up to volume in 25 ml volumetric flask with mobile phase.

Calibration curve (general procedure)

Calibration curves were prepared by taking appropriate aliquots of the standard stock solutions of the drug in different 10 ml volumetric flasks and the solutions were made up to volume with phosphate buffer pH 3.5 to give final concentrations of 8-120 μ g/ml and 10-100 μ g/ml for VAL and LOS, respectively. These standard solutions were analyzed in five replicates. The peak areas were plotted against concentration and the data was subjected to linear regression. Fig. 2a and Fig. 3a show standard chromatograms of 25 μ g/ml valsartan and losartan, respectively.

Preparation of marketed formulations for assay

Ten tablets were weighed, finely powdered (or the content of 10 capsules for valsartan), and an accurately weighed sample of powdered tablets or capsules equivalent to 25 mg of losartan or valsartan was treated with few milliliters of mobile phase. The solution was then sonicated for 30 minutes, this solution was filtered through 0.45 μ m filter paper and then diluted with mobile phase in a 25 ml volumetric flask to yield a starting concentration of 1 mg/ml.

Forced degradation studies

In order to determine whether the analytical method and assay were stability-indicating, valsartan and losartan active pharmaceutical ingredient (API) powder were stressed under various conditions to conduct forced degradation studies.

Acid, alkaline and neutral degradation

Forced degradation in acidic and alkaline media was performed by adding 20 ml of either HCl or NaOH (0.1, 0.5 or 1N) to 10 ml of the standard stock solution (1 mg/ml) in a 100 ml tightly capped conical flasks. The flasks were kept in dark in a water bath thermostated to 80° C up to 3h and then neutralized using NaOH or HCl of appropriate concentration then transferred to 100 - volumetric flasks and diluted to volume with phosphate buffer pH 3.5. Diluted solutions were prepared by appropriate dilution in the same solvent. Degradation under neutral condition was performed by adding 20 ml of distilled water instead of acid or base.

Oxidative degradation

20 ml of either 3.0 or 30.0 % hydrogen peroxide was added to 10 ml of standard stock solution then kept in dark at 40 ° C in a thermostated water bath up to 24 h. The solution was then completed to volume with phosphate buffer pH 3.5 in a100 ml volumetric flask.

Thermal degradation

Thermal treatment was performed by exposing the drug to a controlled temperature oven at 60 °C for three days in tightly closed vials. The solution was then prepared in phosphate buffer pH 3.5 to achieve final concentration 100 μ g/ml. The proposed HPLC method (general procedure) was then applied to the drugs kept under this condition and as well as under the same conditions but protected from light and heat (at room temperature, 35±2 °C and 75±5 % RH).

Stability on exposure to air

Effect of air was performed by keeping each of the drugs in Petri plates in a thin layer of 1mm thickness in dark, at room temperature (35±2 °C) and 75±5 % RH for one week. The proposed HPLC method was applied to the drugs kept under these conditions as well as under the same conditions but protected from air,

RESULTS AND DISCUSSION

HPLC method: Development and optimization

Method development and optimization

An isocratic method was found suitable to optimize the separation of major degradation products formed under various stress conditions. In the preparative chromatogram steps, UV overlain spectra of the two drugs and of all degradation solutions showed that valsartan, losartan and their degradation solutions absorbed appreciably at 225 nm. So, detection at this wavelength was selected for method development. The composition, pH, flow rate of the mobile phase and column temperature were changed to optimize the separation conditions. A mobile phase consisting of 0.02 mM NaH₂ PO₄ buffer (pH 3.5) and acetonitrile (40:60 v/v), a flow rate of 1.5 ml/min and column temperature maintained at

40 °C was found to yield satisfactory retention time with sharp symmetrical peaks. The standard chromatogram of each drug (50 μ g/ml) is shown in Fig. 2 a and Fig. 3 a.

Results of forced degradation studies

Forced degradation studies were performed for bulk drug, to provide an indication of the stability indicating property and specificity of the proposed method. This degradation is mainly observed in terms of loss of assay.

Acid and alkaline degradation

By keeping standard solution of valsartan or Iosartan in 0.1N, 0.5N and 1N at 80 ° C up to 3h, about 12 or 8 % of valsartan or losartan was degraded in 0.1N HCI. Increasing the HCI valsartan showed a higher concentration sensitivity to the acid than losartan; since in 0.5 N and 1N HCl, VAL concentration decreased of 40 and about 68%, respectively, with appearance of three new additional peaks at 1.46, 1.81 and 4.45 min beside the drug peak at about 3 min: Losartan concentration decreased 15 and 19 % respectively; with appearance of only a small peak at retention time 1.46 min (Fig. 2b & Fig. 3b) in addition to the drug peak at about 2.6 min. At more drastic condition, when heating the drug with 2N HCl at 100 °C for 8 h, losartan lost about 28% of its concentration while valsartan lost about 78% of its concentration, which shows the great sensitivity of valsartan to HCI acid compared to losartan. When heated with 0.1N NaOH at 80 °C for 3 h no sufficient degradation was observed for both drugs. Treatment with 0.5 N NaOH at 80 °C for 3 h. 88% and 72% of losartan and valsartan remained, respectively; with appearance of only a new small peak at about 1.470 min for both drugs in addition to smaller one at 2.48 min for By using 1N NaOH further valsartan. degradation was observed for valsartan more than losartan (Fig.2c and Fig.3c).

Oxidative degradation

When valsartan or losartan standard solution was kept in 3% hydrogen peroxide up to 24 h at 40 °C no degradation was observed for valsartan while losartan degraded by about 6%. In 30% H_2O_2 when kept at 40° C for up 24 h, losartan showed great sensitivity, it lost about 94% of its assay with appearance of many additional small peaks around the main peak of the drug while valsartan showed about 14 % decrease in its concentration (Fig.2d and Fig.3d). This result confirms the oxidizable nature of losartan which was confirmed by the previous work done by Zhao et al. [20] in which two main losartan oxidative degradates were identified when losartan tablets were stored for three years at 40 °C /75% RH. They were formed by oxidation of hydroxyl group present in losartan (-CH-OH) to form aldehyde form(-CHO) or keto degradate (-C=O). Also, M.,Vidyawathi et al. in a pharmacokinetic study and due to presence of the hydroxyl group, they showed the tendency of losartan to form two oxidation degradates or active metabolites using fermentation techniques(carboxylic acid and 3'- hydroxy losartan metabolites) [29].

Stability in dissolving solvents and on exposure to air.

Losartan and valsartan were found to be nearly stable when kept in water at 80°C for 3h, less than 2 % degradation was observed and when standard stock solution of valsartan in methanol or losartan in water was kept at room temperature ($35\pm2°C$) for one weak, losartan showed 1.2 % decrease and the peak area of valsartan in methanol diminished by about 5%.

On exposure to open air, in dark at room temperature $(35\pm2\,^{\circ}C)$ and $75\pm5\,^{\circ}$ RH for one week, losartan concentration showed a decrease of about 4% while valsartan remained stable.

Thermal stability

Thermal stability of the two drugs was studied by exposing the drug to hot air (in oven at 60 °C) for three days. The proposed HPLC method (general procedure) was then applied to the drugs kept under this condition and as well as under the same conditions but protected from heat (ambient temperature). Results showed a decrease in valsartan concentration of about 5% and no change was observed for losartan (stress degradation results are summarized in table 1). In all cases, the proposed HPLC method effectively separates the drugs from their degradation products and thus can be used as stability-indicating.

Validation of the method (analytical parameters)

The method of analysis was validated as per the recommendations of ICH [<u>30</u>] for the parameters like linearity, Intra and inter-day precision, ruggedness, robustness, accuracy, limits of detection (LOD) and quantification (LOQ), and specificity.

Linearity

Linearity was established by least squares linear regression analysis of the calibration curve. Peak areas of valsartan and losartan were plotted against their respective concentrations and linear regression analysis performed on the resultant curve. The constructed calibration curves were linear over the concentration range of $8-120\mu$ g/ml and $10-100\mu$ g/ml for VAL and LOS, respectively. Correlation coefficient (*n*=3) was found to be 0.999 and 0.998 for VAL and LOS, respectively. Regression line equation was found to be:

Y= -65.25+52.92 X and Y= 69.68+ 64.04 X for valsartan and losartan, respectively.

The limit of detection (LOD) and limit of quantitation (LOQ)

The limit of detection (LOD) and limit of quantitation (LOQ) were calculated using following formulae: LOD= 3.3(SD)/S and LOQ= 10 (SD)/S, where SD=standard deviation of response (peak area) and S= average of the slope of the calibration curve. LOD values for valsartan and losartan were found 2.11 µg/ml and 2.99 µg/ml, respectively, and limits of quantification were found 6.40 µg/ml and 9.07 µg/ml respectively.

Precision

Intra-day precision of the method was studied by calculating the relative standard deviation (RSD %) of the peak area for six determinations of solutions of pure drug at three concentrations 20, 50, and 80 µg/ml. All determinations were made on the same day and under the same experimental conditions. Inter-day precision was assessed by injecting the same three concentrations over three consecutive days, results (Table 2) indicate acceptable level of inter-day and intra- day precision. The ruggedness of the method was assessed by comparison of the intra- and inter-day assay results for valsartan or losartan undertaken by two analysts. The % RSD values for intra- and inter-day assays of the drug performed in the same laboratory by the two analysts did not exceed more than 2%, thus indicating the ruggedness of the method.

Recovery (standard addition method)

Recovery studies by the standard addition method were performed. For this reason, a known amount of the pure drug (50 μ g) which was previously analyzed, were spiked with 30, 50 or 75 μ g losartan or valsartan standard solutions, and the mixtures prepared in triplicate, were analyzed. The values of recovery % and RSD % listed in table 3 indicate that the method is accurate.

Specificity

The results from the stress studies indicated that the method was highly specific for VAL and LOS. The degradation products were completely distinguishable from the parent compound. Based on peak purity of VAL and LOS, every degradation sample showed that the peaks were homogenous and no co eluting peaks were observed indicating that the method was stability indicating and specific.

ASSAY

The validated method was applied to the determination of valsartan and losartan in commercially available disartan capsules and kanzar tablets. The results of the assay (n = 5) undertaken yielded 99.73% and 100.40 (% RSD less than 2%) of label claim for valsartan and losartan, respectively (Table 4). The results of the assay indicate that the method is selective for the analysis of valsartan or losartan without interference from the excipients used to formulate and produce these capsules or tablets.

CONCLUSION

A simple, rapid, accurate, and precise stabilityindicating HPLC analytical method has been developed and validated for the routine analysis of valsartan or losartan in their formulations without any interference from excipients. The results of stress testing reveal that the method is selective and stability indicating. The proposed method has the ability to separate the drug from their degradation products, and can be applied to the analysis of samples obtained during accelerated stability experiments. Results showed that LOS is highly susceptible to oxidation. Consequently, adequate air protection should be adopted for its packaging, storage and handling; while VAL is very sensitive to acid hydrolysis and exhibits higher sensitivity to alkaline medium than LOS. The two drugs showed nearly the same stability under neutral hvdrolvsis.



Fig. 2a: Chromatogram of Valsartan standard 25 µg/ml



Fig. 2b: Chromatogram of Valsartan degradation product 25 μg/ml in 1N HCI



Fig. 2c: Chromatogram of Valsartan degradation product 25 µg/ml in 1N NaOH



Fig. 2d: Chromatogram of Valsartan degradation product 25 μ g/ml in 30% H₂O₂



Fig. 3a: Chromatogram of Losartan standard 25 µg/ml



Fig. 3b: Los Chromatogram of artan degradation product 25 $\mu g/ml$ in 1N HCl



Fig. 3c: Chromatogram of Losartan degradation product 25 $\mu g/ml$ in 1N NaOH



Fig. 3d: Chromatogram of Losartan degradation product 25 $\mu g/ml$ in 30% H_2O_2

and losar tan Using Different conditions				
	Exposure	Heating	Assay %	
Agent	time(h)	temperature (°C)	valsartan	losartan
		HCI		
0.1N	3	80	88.0	92.0
0.5N	3	80	60.0	85.0
1N	3	80	32.3	81.4
2N	8	80	22.0	72.0
NaOH				
0.1N	3	80	96.0	98.0
0.5N	3	80	72.0	88.0
1N	3	80	59.4	87.0
H ₂ O ₂				
3%	24	40	99.3	94.1
30%	24	40	86.1	6.2
Solvents				
H ₂ O	3	80	98.7	98.0
	7 days	Room temp 35±2	-	98.8
Methanol	7 days	Room temp 35±2	95.0	-
Open air 75± 5 % RH	7 days	Room temp 35±2	99.8	95.9
oven	3 days	60	95.3	99.9

Table 1: Results of the Stress Degradation Tests of valsartan and Iosartan Using Different Conditions

Table 2: Intra-day and inter-day precision

Conc (µg/ml)	Intra-day precision Mean area± SD RSD %		inter-day precision Mean area± SD RSD %	
	VAL	LOS	VAL	LOS
30	29.87± 0.33	29.98±0.40	29.93±0.49	29.82±0.35
	1.10	1.33	1.64	1.17
50	50.14±0.45	50.29±0.63	49.98±0.73	50.12±0.54
	0.90	1.25	1.46	1.08
80	80.16±0.87	80.12±0.50	79.87±1.11	80.19±0.67
	1.08	0.62	1.39	0.83

Table 3: Results from recovery studies with valsartan
and losartan by standard addition method

drug	Initial amount (μg/ml)	Amount added (μg/ml)	Amount recovered (μg/ml) mean± SD	RSD %	Standard analytical error (SAE)
Valsartan	50	30	29.99±0.41	1.36	0.24
		50	50.12±0.32	0.64	0.18
		75	74.95±0.51	0.68	0.29
Losartan	50	30	30.02±0.44	0.145	0.25
		50	49.96±0.25	0.5	0.15
		75	75.10±0.34	0.45	0.2

Table 4: Assay results for determination of valsartan and losartan in pharmaceutical formulations

Generic name	Brand	Recovery(%) ± SD* RSD %		
	name	Proposed method	Reported method	
Valsartan	Disartan capsules	99.73 ± 0.29	99.51 ±0. 63	
	80mg/capsule	0.29	0.64	
Losartan	Kanzar tablets	100.40 ± 0.64	99.87±0.66	
	100mg/tablet	0.63	0.66	

REFERENCES

- 1. Sweetman Sc. Martindale. The extra Pharmacopeoia. The Complete Drug Reference: Pharmaceutical Press, London, 2009; 36 th Edition.
- 2. United States Pharmacopeia, US Pharmacopeial Convention. Rockville MD 2012; 35 NF30.
- 3. Rahul R Nahire, Sagar S Joshi, Varsha Meghnani, Nalini Shastri, Surendra KV Nath and Sathish J. Stability indicating RP-HPLC method for simultaneous determination of amlodipine besylate and valsartan combination in bulk and commercial dosage forms. Asian Journal of Pharmacy and Life Science. 2012; 2 (2): 280.
- 4. Bhaskara Raju V and Lakshmana Rao A. Reversed phase HPLC analysis of valsartan in pharmaceutical dosage forms. International Journal of Chemical Environmental and Pharmaceutical Research. 2011;2(1):56-60.
- Akiful Haque M, Hasan S Amrohi, Prashanth Kumar K, Nivedita G, Pradeep Kumar T, Dibyalochan Mohanty and Prakash V Diwan. Stability indicating RP-HPLC method for the estimation of valsartan in pharmaceutical dosage form. IOSR Journal of Pharmacy. 2012;2 (4):12-18.
- Rao KS, Jena N and Rao M. Development and validation of a specific stability indicating high performance liquid chromatographic method for valsartan. J Young Pharmacists. 2010; 2(2):183-189.
- Shah NJ, Suhagia BN, Shah RR and Patel NM. HPTLC method for the simultaneous estimation of valsartan and hydrochlorothiazide in tablet dosage Form. Indian Journal of Pharmaceutical Sciences. 2009;71(1):72-74.
- Yan J, Wang W, Chen L and Chen S. Electrochemical behavior of valsartan and its determination in Capsule. Colloids Surf B Biointerfaces. 2008;67(2):205-209.
- Hillaert S and Van den Bossche W. Optimization and validation of a capillary -zone electrophoretic method for the analysis of several angiotensin-II-receptor antagonist. Journal of Chromatography A. 2002;979(1-2):323-333.
- 10. Stolarczyk M, masalanka A, Krzek J and Milczarek J. Application of derivative spectrophotometry for determination

of enalapril, hydrochlorothiazide and valsartan in complex pharmaceutical preparations. Acta Poloniae Pharm Drug Res. 2008;65(3):275–281.

- 11. Tatar S and Saglik S. Comparison of UVand second derivative spectrophotomtric and LC methods for the determination of valsartan in pharmaceutical formulation. Journal of Pharmaceutical and Biomedical Analysis. 2002;30:371–375.
- 12. Kokila Parmar and Jignesh Shah. Simultaneous estimation of aliskiren and valsartan by ratio spectra derivative spectrophotometry method in their fixed dosage forms. International Journal of ChemTech Research. 2014;6(2):1268-1275.
- 13. United States Pharmacopeia, US Pharmacopeial Convention. Rockville MD 2012; 35 NF30.
- 14. Rao KS and Srinivas K. RP-HPLC method for the determination of losartan potassium and ramipril in combined dosage Form. Indian J Pharm Sci. 2010;72(1):108-11.
- 15. Bonfilio R, Tarley CR, Pereira GR, Salgado HR and de Araújo MB. Multivariate optimization and validation of an analytical methodology by RP-HPLC for the determination of losartan potassium in capsules. Talanta. 2009;80(1):236-41.
- 16. Sivakumar T, Venkatesan P, Manavalan R and Valliappan K. Development of a HPLC method for the simultaneous determination of losartan potassium and atenolol in tablets. Indian Journal of Pharmaceutical Sciences. 2007;69(1):154-157.
- Baing MM, Vaidya VV, Sane RT, Menon SN and Dalvi K. Simultaneous RP-LC determination of losartan potassium, ramipril and hydrochlorothiazide in pharmaceutical preparations. Chromatographia. 2006;64(5):293 – 296.
- 18. Ulu ST and Saglik S. Comparison of UV and Second derivative spectrophotometric and HPLC methods for the determination of Losartan in tablets. Turk J Pharm Sci. 2004;1:165-75.
- Deanne L Hertzog, Jennifer Finnegan McCafferty, Xueguang Fang R Jeffrey Tyrrell and Robert A Reed. Development and validation of a stability-indicating HPLC method for the simultaneous determination of

losartan potassium, hydrochlorothiazide, and their degradation products. Journal of Pharmaceutical and Biomedical Analysis. 2002;30(3):747-760.

- Zhao Z, Wang Q, Tsai EW, Qin XZ and Ip D. Identification of Iosartan degradates in stressed tablets by LC-MS and LC-MS/MS. J Pharm Biomed Anal. 1999;20(1-2):129-36.
- 21. McCarthy KE, Wang Q, Tsai EW, Gilbert RE, Ip DP and Brooks MA. Determination of Iosartan and its degradates in COZAAR tablets by reversed-phase high-performance thinlayer chromatography. J Pharm Biomed Anal. 1998;17(4-5): 671-7.
- 22. Santhana Lakshmi K and Lakshmi S doi. Simultaneous analysis of Iosartan potassium, amlodipine besylate and hydrochlorothiazide in bulk and in tablets by high-performance thin layer Chromatography with UV-Absorption Densitometry. J Anal Methods Chem. 2012; 2012:108281.
- 23. Balesteros Manoela R; Faria Adriana F and Oliveira Marcone ALde. Determination of Iosartan associated with chlorthalidone or hydrochlorothiazide in capsules by capillary zone electrophoresis. J Braz Chem Soc. 2007;18(3): 554-558.
- 24. Hillaert S and Van den Bossche W. Optimization and validation of a capillary zone electrophoretic method for the analysis of several angiotensin-II-receptor antagonists. J Chromatogr A. 2002;979(1-2):323-333.
- 25. Swetha Y, Malleshwari P, Jyotsna Dr. PRamalingam, Hari D Hara Theja and

Vinod kumar K. New UV-Spectrophotometric method for the determination of losartan potassium in pharmaceutical dosage form and its application to protein binding study. Journal of Pharmacy Research. 2011;4(11):4139-4141.

- Soad S Abd El-Hay, Magda Y El-Mammli and Abdalla A Shalaby. Determination of clemastine hydrogen fumarate, desloratadine, losartan potassium and moxepril HCl through binary complex formation with eosin. Arabian Journal of Chemistry. 2011; 10.1016/j.arabjc.2011.06.021.
- 27. Ibrahim A Darwish. Analytical study for the charge-transfer complexes of losartan potassium Analytica Chimica Acta. 2005; 549(1-2):212-220.
- 28. Ulu-ST and Saglik S. Comparison of UVand second derivative spectrophotometric and highperformance liquid chromatographic methods for the determination of losartan in tablets. Turk-J-Pharm-Sci. 2004;1(3):165-175.
- 29. Vidyawathi M, Krishna DR, Prasad KVS and Vidyasagar. Studies on metabolism of losartan using microbes, International journal of pharmaceutical sciences and nanotechnology. 2008; 1(1): 52-59.
- ICH Harmonized Tripartite Guideline, Validation of Analytical Procedures Text and Methodology, O2(R1) Current Step 4 Version, Parent Guidelines on Methodology Dated November 1996, Incorporated in November 2005.